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COMPARATIVE ASSESSMENT OF SOIL PHYSICO-CHEMICAL PROPERTIES IN THE ECOLOGICAL ZONES OF BAYELSA STATE, NIGERIA

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ABSTRACT

The study examined the comparative assessment of soil physico-chemical properties in the ecological zones of Bayelsa State, Nigeria. Twenty 20m x 20m quadrats were delineated in the natural vegetation of the freshwater swamp (FWS) and mangrove (MG) ecological zones to collect soil samples and tree species. The soil samples were collected from the soil depths from 0-15cm, 15-30cm and 30-45cm and were analysed in the laboratory with standard methods. Descriptive and inferential statistics were used for data analysis. Findings showed that particle size composition (sand , silt, clay) were significantly higher in the freshwater than the mangrove. Similarly, soil temperature was significantly higher in the freshwater and mangrove. It is recommended that more soil studies should be carried out in the study area and moreso, the bulk density and temperature should be closely and adequately monitored for the adequate retention of soil fertility for the survival of plants.

Keywords: Physico-chemical properties, Freshwater, Mangrove, Particle size composition, Bayelsa State.

1. INTRODUCTION

Soils have become one of the most endangered natural resources in the world under current pressures from land degradation and climate change (Titeux et al., 2016). Each year, an estimated 25-40 billion tons of fertile soil are lost globally (FAO and ITPS, 2015). Hence, improving soil health through sustainable land management should be a common goal for farmers and land managers, to protect, maintain and build soil which is their most vital resource. Soils are the major reservoir of C in terrestrial ecosystems, and soil C plays a vibrant role in influencing the global C cycle and climate change while regulating soil health and productivity (Mehra, Singh, Kunhikti and Cowie, 2018; Singh, Setia, Wiesmeier and Kunhikrishnan, 2018). Soil health thus refers to the capacity of the soil to perform a range of agronomic and ecosystem functions, in order to sustain biological productivity, maintain environmental quality and promote plant and animal health (Kibblewhite, Ritz, and Swift, 2008; Lal, 2011; Mehra et al., 2018). Soils that are healthy occur when their biological, chemical, and physical characteristics are all adequate and are able to enable high yields of crops. As this occurs, roots are able to proliferate easily, bountiful water enters and stored in the soil, the plant has a sufficient nutrient supply, there are no harmful chemicals in the soil, and beneficial organisms are very active and able to keep potentially harmful ones in check as well as stimulate plant growth. Furthermore, soil physical health is largely determined by the impact of management and climate factors on

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SOM, soil structure, bulk density and air and water movement through the soil profile (Luo et al., 2017). One of the most important attributes of a healthy soil is soil structural stability, which affects the movement and management of water, oxygen and nutrients (Datiri and Lowery, 1991; Ussiri et al., 2009; Kladivko, 2001).

Thus, changes in soil structure which result directly from anthropogenic activities and indirectly from climate change can adversely influence soil physical health (Luo et al., 2017). Studies have demonstrated that poor soil structure due to anthropogenic activities such as continuous cultivation inhibits many plant processes (germination, root growth) (Turmel et al., 2015). Moreover, a boost in air and soil temperature unavoidably creates disparity in the soil-thermal regime, which is governed by the processes like evaporation, heat conduction and convective transfer via the movement of gas and water through the soil profile. This imbalance of soilthermal regimes can impact important soil processes such as rapid decomposition of SOM and nutrient loss via volatilization (Qian et al., 2011). It is believed that the biological effectiveness proves that below ground organisms are significant in determining the functioning of ecosystems as they substantially regulate decomposition or nutrient mineralization/cycling. Hence, they partly determine plant growth and sustain the long-term productivity (Wolters, 2001; Rossi, Mathieu, Cooper and Grimaldi, 2006). As a result, soil organisms are increasingly considered as a resource to be managed and protected (Rossi et al., 2006). Soil is not a state factor for plant growth, as soil formation is influenced by plants (biota) climate, geology, topography and time (Fujii et al., 2018). Soil can influence plant physiological processes, while plants can also change soil processes, a concept known as plant-soil feedback. The cause-effect in plant-soil processes or plant-soil feedback are linked to soil chronosequences, plant impacts on soil processes, strategies of nutrient acquisition and utilization (allocation), and niche differentiation related to soil nutrients.

Soil is known for its huge spatial (vertical and even) and transient heterogeneity which leads to a wide scope of surface kinds, total and pore estimates and microclimates, and a scope of assets and asset apportioning in reality. This intricacy is a snag to the utilization of single measures (for example pH, natural matter substance) as comprehensively pertinent signs of soil wellbeing and environment work (Baveye et al. 2016), rather basic markers should be developing or possibly very much connected to basic systems. The actual climate can be considered as a format on which living beings and biological frameworks work; for some dirt living beings, particularly miniature organic entities, the design of the dirt pore network de-fines the powerful territory space in soil (Young and Ritz, 2000). The sum and nature of the pore space in soil are reliant on soil surface as well as on the collection of mineral particles and soil natural matter (SOM), which is, the development and adjustment of soil structure. Most soil organic entities have restricted movement limit (Fitter et al. 2005) and motility of many soil species is low contrasted with the size of asset inconsistency (Ettema and Wardle, 2002). Soil living beings likewise regularly enter inert or torpid states in negative circumstances, with the goal that variety is safeguarded much under outrageous circumstances; this is undifferentiated from the job of soil seed-banks in protecting plant variety (Ettema and Wardle, 2002). Henceforth, creatures' reaction to the actual climate might show designs that change among species and are obliged by the math of the climate (Williams, Marsh and Winter, 2002).

A few investigations have been done at various spatial and transient scales for recognizing the vegetative elements as well as the connections among various vegetation types and critical

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changes at worldwide and local level have been seen in environment as well as in vegetation cover. Under such circumstances, more consideration has been paid throughout the most recent a long time on the environment related investigations. Nonetheless, it is obvious that the vegetation is firmly interrelated with their general surroundings. Changes in various climatic boundaries could result to the arrangement and variety of vegetation cover at different levels. They can be extreme and surprisingly unsafe on certain types of vegetations and can be minor on a few others. Thus, an equivalent consideration must be paid on the vegetation elements. All the more critically the connections that could exist among these interrelated parts must be recognized to make precise and sensible expectations on the changing states of the vegetation as well as the climatic boundaries. This will likewise give the chance to making prudent steps for limiting the potential dangers related with such changing states of environment as well as the biological systems. It is likewise expected that the provincial varieties of various climatic boundaries to be a lot bigger and consequently nitty gritty examinations on explicit issues must be done to get an unmistakable picture on these progressions at territorial scales. Current interests in the livability of the earth and climate have animated a large group of worldwide observing projects to analyze the potential results of varieties in prime environment boundaries. for example, precipitation, temperature barometrical gas focus, radiation levels, and land cover.

2. MATERIALS AND METHODS

The study was carried out in Bayelsa State, Nigeria. Bayelsa State is geographically located with latitude 4°15'North and latitude 5°23'South and longitude 05°22' west and 06°45' East. It shares boundaries with Delta State on the North, Rivers State on the East and the Atlantic Ocean on the West and South. The State has a land mass of 21, 110 Sq km/10,773 square miles. Bayelsa is a State in Southern Nigeria in the core Niger Delta region. The area lies almost entirely below sea level with a maze of meandering creeks and rivers in the south, all flow into the Atlantic ocean via the major rivers such as San Bartholomew, Brass, Nun, Ramos, Santa Barbara, St Nicholas, Sangana, Fishtown, Ikebiri creek, Middleton, Digatoro creek, Penington and Dobo (Figure 1). The study area has the same weather condition like Port Harcourt as a region. The climatic condition is the tropical climate (Ologunorisa & Adejuwon, 2003). The months of February and March records the highest temperature of 33°C, while the months of January and December records the lowest temperature of 21°C. Temperature rises through the months of October, November and December. The monthly rainfall in the area is slightly predictable due to climate change in the world today, temporally rainfall decreases from the months of October to February which is the dry season (Ologunorisa and Adejuwon, 2003). The rainfall in the area is greatly influenced by the Inter Tropical Discontinuity (ITD), due to its location. Rainfall peaks in July and September with a little dry season in August (August break). The area experiences about 2476 mm of precipitation falls annually. There are three major soil groups identified in the Niger Delta, namely: the marine and fluvial marine sediments; the mangrove swamp alluvial soils; and freshwater brown loams and sandy loams. The "upland" area was originally occupied by rainforest which has been drastically modified by human activities. In most places, economic trees, particularly oil palm, have been preserved and thus the sobriquet for this vegetation as "oil palm bush." The riverine area is divided into three main hydro vegetation zones namely, the beach ridge zone, the saltwater zone and the freshwater zone. In terms of general surface features, the area is very unique, and falls within the coastal belt dominated by low lying coastal

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plain which belongs to the structural sedimentary formation of the recent Niger Delta. Umeuduji and Aisebeogun (1999) identified that the area is within the belt of fresh water raffia dominated wetlands with heights which vary between 10-25m above sea level. Five soil samples were collected from each 20m x 20m quadrat using soil auger at the depth of 0-15cm, and 15-30cm and 45-60cm. The soil samples in each depth were bulked together into a plastic container and a composite soil sample was taken in each quadrat from topsoil and subsoil. Thus, 20 soil samples were collected from each 20m x 20m quadrat the depth of 0-15cm, 15-30cm and 45-60cm in each ecological zone. Composite soil samples were collected into well-labelled polythene bags and brought into the laboratory. The soil samples were air-dried and carefully sieved with 2mm diameter mesh in order to separate the soil from stones. Soil temperature was measured with soil thermometer in situ (Ochsner, 2008) while soil moisture was measured using gravimetric method (Su, Singh & Baghini, 2014). Soil pH was determined using saturated paste extract while organic carbon was determined by Walkey and Black's rapid titration method (Walkey & Black, 1934). Descriptive statistics was used to describe the mean values of the soil properties and pairwise t test was used to determine the significant variation in the soil properties between freshwater swamp and mangrove.



Figure 1: Ecological Zones of Bayelsa State

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3. RESULTS AND DISCUSSIONS

Soil Physical and Chemical Properties in the Ecological Zones

The physical properties of soil at varying soil depths are shown in Table 1 while Table 2 shows the general descriptive statistics of soil physical properties. Sand content was highest in FWS zone (50.68%) at soil depth of 15-30cm and the lowest was observed also under the FWS zone (31.61%). Considering silt content, FWS zone recorded 52.99% (0-15cm), while M zone recorded 32.25% (0-15); FWS recorded 34.36% (15-30cm) while M zone recorded (32.59% (15-30cm); and 45.41% (30-45cm) under the FWS zone while M zone recorded 30.54% (30-45cm). Clay content was highest (28.38%) at soil depth of 30-45cm under the M zone and lowest (14.33%) at soil depth of 0-15cm under the FWS zone. The bulk density slightly varied in the sampled ecological zones (FWS and M), but FWS zone recorded the highest with mean value of 1.58 g/cm³ at soil depth of 15-30cm and the least was observed in M zone (1.514 g/cm³) at soil depth of 30-45cm. The porosity (%) and water holding capacity (%) varied slightly among the sampled ecological zones. However, soil porosity was highest in the M zone (47.47%). This may be attributed to the high sand content which might have enabled wider pore space within the soil. The water holding capacity was highest in the M zone with the mean value of 43.27% at soil depth of 30-45cm. Soil moisture was highest in the FWS zone (79.71%) at soil depth of 30-45cm and the lowest was observed in the M zone (72.07%) at soil depth of 15-30cm. Temperature varied considerably in both sampled ecological zones but temperature was highest in FWS zone with a mean temperature of 29.09°C at soil depth of 0-15cm. The general descriptive statistics of soil physical properties on Table 4.97 revealed that among the soil particles size distribution (sand, silt and clay), sand content (45.56%) recorded the highest under the M zone. Overall mean values for Bulk density and water holding capacity showed that they were slightly higher under the FWS zone. Overall mean Porosity (47.20%) was higher under the M zone which was attributed to the higher sand content in soil. Overall mean Temperature $(26.40^{\circ}C)$ and soil moisture content (78.69%) were higher under the FWS zone.

Soil Properties	perties Soil Depth		Mangrove
		Mean±SD	Mean±SD
Sand (%)	0-15	32.68±9.4	46.20±23.8
	15-30	50.68±11.4	49.39±18.9
	30-45	31.61±10.5	41.09±16.8
Silt (%)	0-15	52.99±9.0	33.25±11.6
	15-30	34.36±9.9	32.59±13.3
	30-45	45.41±8.5	30.54±10.7

Table 1: Descriptive Statistics of Soil Physical	Properties across the Soil Depths in Diff	ferent
Ecological Zones		

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Clay (%)	0-15	14.33±5.1	20.56±14.7
	15-30	14.97±4.1	18.02±7.6
	30-45	22.99±9.0	28.38±15.13
Bulk Density (gcm ³)	0-15	1.56±0.1	1.52±0.1
	15-30	1.58±0.1	1.54±0.1
	30-45	1.55±0.1	1.51±0.1
Water Holding Capacity	0-15	42.45±0.7	41.87±0.7
	15-30	42.79±1.1	42.47±1.4
	30-45	43.06±9.0	43.27±1.4
Porosity (%)	0-15	46.52±0.9	47.47±1.5
	15-30	45.99±1.3	46.72±1.6
	30-45	45.91±0.4	47.42±1.5
Soil Moisture (%)	0-15	79.32±3.7	75.07±3.85
	15-30	77.04±4.8	72.07±5.9
	30-45	79.71±4.0	76.47±7.2
Temperature (°C)	0-15	29.09±0.9	26.33±0.8
	15-30	26.12±1.1	24.34±0.6
	30-45	23.98±1.1	22.57±1.4

N=20

Table 2: General Descriptive Statistics of Soil Physical Properties across the Soil Depths in Different Ecological Zones

Soil Properties	Freshwater	Mangrove
	Mean±SD	Mean±SD
Sand (%)	38.32±13.5	45.56±20.1
Silt (%)	44.25±11.3	32.12±11.8

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Clay (%)	17.43±7.5	22.32±13.5
Bulk Density (gcm ³)	1.56±0.1	1.53±0.1
Water Holding Capacity (%)	42.77±1.0	42.54±1.3
Porosity (%)	46.14±0.9	47.20±1.6
Soil Moisture (%)	78.69±4.3	74.54±6.0
Temperature (°C)	26.40±2.4	24.41±1.8

N=60

The information for soil chemical properties across soil depths were displayed on Table 3 while Table 4 reveals the general statistics using their overall mean values in sampled ecological zones. The exchangeable acidity in the sub soil (15-30cm) was highest (0.47 Cmol/kg) in the FWS zone and the lowest (0.40 Cmol/kg) was also experienced under the FWS zone at soil depth of 30-45. The mean highest CEC for FWS zone was 15.16 Cmol/kg at soil depth of 15-30cm while the highest of 15.92 Cmol/kg was experienced under the M zone at soil depth of 30-45cm. The highest mean percentage concentration of organic C was 2.16% (0-15cm) in the FWs zone; while it was 2.12% in M zone at soil depth of 15-30cm. Similarly, total N was highest in the FWS zone (0.38%) at soil depth of 30-45cm and the least was found in M zone with mean value of 0.31% at soil depth of 30-45cm. Available P in the topsoil (0-15cm) was 4.33 mg/kg and the highest of 4.54 mg/kg in deeper subsoil (30-45cm) was also recorded under the FWS zone while the least of 0.77 mg/kg was recorded at subsoil (15-30cm) under the M zone. The mean value of Na varied across soil depths and ecological zones. The highest mean content of Na was 3.15 Cmol/kg at soil depth of 30-45cm and the least content of 0.35 Cmol/kg were recorded under the FWS zone. The exchangeable Ca was highest in the M zone (9.01 Cmol/kg) while the lowest was observed in FWS zone with mean content value of 7.40 Cmol/kg. Exchangeable Mg ranged from 2.72 Cmol/kg (0-15cm) to 4.25 Cmol/kg under the FWS zone. The range for Exchangeable Mg under the M zone was between 3.13 Cmol/kg and 3.61 Cmol/kg. Exchangeable K was highest (0.74 Cmol/kg) at soil depth 0-15cm and lowest at soil depth 30-45cm (1.24 Cmol/kg) under the FWS zone. However, the mean content values of Exchangeable K recorded minimum value of 0.56 Cmol/kg and 1.52 Cmol/kg under the M zone at soil depths of 15-30cm and 0-15cm respectively. In the topsoil (0-15cm), subsoil (15-30cm) and deeper subsoil (30-45cm) the soil pH was acidic across the ecological zones but more acidic (5.45) at soil depth of 30-45cm and also less acidic (5.74) at soil depth of 0-15cm under the M zone. The information for the general descriptive statistics of soil chemical properties across soil depths is displayed on Table 4.

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Table 3. Descriptive Statistics of Soil Physical Properties across the Soil Depths						
Soil Properties	Soil Depth	Freshwater	Mangrove			
		Mean±SD	Mean±SD			
Exchangeable Acidity	0-15	0.45±0.1	0.43±0.1			
(Chiol/Kg)	15-30	0.47±0.1	0.44±0.2			
	30-45	0.40±0.1	0.45±0.1			
Cation Exchange	0-15	11.80±3.6	15.27±8.1			
Capacity (Chiol/kg)	15-30	15.16±6.2	13.11±5.3			
	30-45	13.30±3.8	15.92±11.0			
Organic C (%)	0-15	2.16±0.9	1.92±1.0			
	15-30	1.99±0.7	2.12±1.1			
	30-45	1.79±0.8	1.86±0.9			
Total N (%)	0-15	0.35±0.1	0.32±0.1			
	15-30	0.36±0.1	0.34±0.2			
	30-45	0.38±0.3	0.31±0.2			
Available P (Cmol/kg)	0-15	4.33±1.0	1.34±0.8			
	15-30	4.13±1.0	0.77±0.3			
	30-45	4.54±0.9	0.94±0.9			
Na(Cmol/kg)	0-15	3.15±0.1	2.12±1.6			
	15-30	0.35±0.4	1.18±0.9			
	30-45	0.50±0.3	2.42±2.3			
Ca (Cmol/kg)	0-15	7.42±1.9	8.25±5.3			
	15-30	7.40±1.7	7.80±4.6			
	30-45	8.24±2.1	9.01±0.7			
Mg (Cmol/kg)	0-15	2.72±1.1	3.33±1.9			

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	15-30	3.19±1.0	3.61±1.3
	30-45	4.25±1.8	3.13±1.2
K (Cmol/kg)	0-15	0.74±0.3	1.52±1.7
	15-30	0.79±0.4	0.56±0.4
	30-45	1.24±0.6	1.39±0.8
рН	0-15	5.57±0.2	5.74±0.3
	15-30	5.64±0.2	5.59±1.1
	30-45	5.59±0.4	5.45±0.6

N=20

Table 4: General Descriptive Statistics of Soil Physical Properties across the Soil Depths

Soil Properties	Freshwater	Mangrove
	Mean±SD	Mean±SD
Exchangeable Acidity (Cmol/kg)	0.44±0.1	0.43±0.1
Cation Exchange Capacity (Cmol/kg)	13.42±4.8	14.77±8.4
Organic C	1.98±0.8	1.97±1.0
Total N	0.36±0.2	0.32±0.1
Available P (Cmol/kg)	4.33±1.0	1.01±0.7
Na (Cmol/kg)	1.42±4.6	1.90±1.8
Ca (Cmol/kg)	7.69±1.9	8.35±5.7
Mg (Cmol/kg)	3.39±1.5	3.36±1.5
K (Cmol/kg)	0.93±0.5	1.16±15
рН	5.60±0.3	5.59±0.5

N=60

Variation in the soil physical and chemical properties among the soil depths and between the ecological zones

The computed ANOVA analysis for soil physical properties under the FWS zone is displayed on Table 5. The results showed that soil physical parameters among soil depths for water holding

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capacity (%) (F=1.897; p<0.05), bulk density (g/cm³) (F=0.323; p<0.05), porosity (%) and soil moisture content (%) (F=2.393; p<0.05) were all not significant. However, there exist significant variations of soil physical parameters among soil depths for sand content (%) (F=20.979; p<0.05), silt content (%) (F=20.846; p<0.05), clay content (%) (F=11.134; p<0.05) and temperature (F=120.114; p<0.05).

The ANOVA statistics of soil chemical properties among soil depths in the FWS zone is displayed on Table 6. The results showed that variations in soil chemical properties among soil depths for Ex. Acidity (F=1.494; p<0.05), Na (F=2.163; p<0.05), CEC (F=2.558; p<0.05), pH (F=0.352; p<0.05), Organic C (F=1.080; p<0.05), Total N (F=0.208; p<0.05), Avail. P (F=0.784; p<0.05), and Ca (F=1.244; p<0.05) were not significant. However, variations in soil chemical properties among soil depths for Mg (F=6.346; p<0.05) and K (F=7.352; p<0.05) were significant under the FWS zone.

 Table 5: ANOVA of Physical Soil Properties among the soil depths in Freshwater

 ecological zone

Soil Parameters		Sum of Squares	df	Mean Square	F	Sig.
Water Holding Capacity	Between Groups	3.737	2	1.869	1.897	0.159
	Within Groups	56.136	57	.985		
	Total	59.873	59			
Sand	Between Groups	4590.937	2	2295.468	20.979	0.000*
	Within Groups	6236.799	57	109.418		
	Total	10827.736	59			
Silt	Between Groups	3512.653	2	1756.326	20.846	0.000*
	Within Groups	4802.317	57	84.251		
	Total	8314.970	59			
Clay	Between Groups	931.504	2	465.752	11.134	0.000*
	Within	2384.342	57	41.831		

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	Groups					
	Total	3315.846	59			
Bulk Density	Between Groups	.010	2	.005	.323	0.725
	Within Groups	.853	57	.015		
	Total	.863	59			
Porosity	Between Groups	4.396	2	2.198	2.430	0.097
	Within Groups	51.548	57	.904		
	Total	55.944	59			
Soil moisture	Between Groups	83.196	2	41.598	2.393	0.100
	Within Groups	990.758	57	17.382		
	Total	1073.954	59			
Temperature	Between Groups	263.473	2	131.737	120.114	0.000*
	Within Groups	62.516	57	1.097		
	Total	325.989	59			

N=60

*Significant at p<0.05

Table 6: ANOVA of Chemical Soil Properties among the soil depths in Freshwater ecological zone

Soil Parameters		Sum of Squares	df	Mean Square	F	Sig.
Ex. Acidity	Between	.052	2	.026	1.494	.233

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	Groups					
	Within Groups	.992	57	.017		
	Total	1.044	59			
Na	Between Groups	89.501	2	44.750	2.163	.124
	Within Groups	1179.080	57	20.686		
	Total	1268.581	59			
CEC	Between Groups	113.322	2	56.661	2.558	.086
	Within Groups	1262.667	57	22.152		
	Total	1375.990	59			
	Within Groups	62.516	57	1.097		
	Total	325.989	59			
рН	Between Groups	.056	2	.028	.352	.705
	Within Groups	4.514	57	.079		
	Total	4.570	59			
Organic C	Between Groups	1.342	2	.671	1.080	.347
	Within Groups	35.422	57	.621		
	Total	36.763	59			
Total N	Between Groups	.021	2	.011	.208	.813
						1

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	Within Groups	2.928	57	.051		
	Total	2.949	59			
Avail. P	Between Groups	1.633	2	.817	.784	.461
	Within Groups	59.364	57	1.041		
	Total	60.997	59			
Са	Between Groups	9.170	2	4.585	1.244	.296
	Within Groups	210.073	57	3.685		
	Total	219.242	59			
Mg	Between Groups	24.461	2	12.231	6.346	.003*
	Within Groups	109.857	57	1.927		
	Total	134.319	59			
K	Between Groups	3.093	2	1.547	7.352	.001*
	Within Groups	11.990	57	.210		
	Total	15.083	59			

N=60

4.CONCLUSION AND RECOMMENDATIONS

The study concluded that the soil physico-chemical properties such as sand content, silt content, clay content, soil temperature, Mg and K varied significantly between freshwater and mangrove ecological zones. Based on the findings, the study recommended that more soil studies are required in between the freshwater and mangrove ecological zones. Moreso, the bulk density and temperature that were higher in the freshwater should be monitored adequately for the purpose of retention of soil fertility.

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REFERENCES

- Aanen D.K., Henrik H., Debets A.J., Kerstes N.A., Hoekstra R.F., and Boomsma J.J. (2009): High symbiont relatedness stabilizes mutualistic cooperation in fungus-growing termites. Science 326:1103–1106
- Abiven S., Menasseri S. and Chenu C., (2009). The effects of organic inputs over time on soil aggregate stability A literature analysis. *Soil Biol. Biochem.*, 41(1):1-12.
- Adejuwon J.O. (2012). Rainfall seasonality in the Niger Delta Belt, Nigeria. Journal of Geography and Regional Planning, 5(2):51-60.
- Adekunle V.A.J., Alo A.A. and Adekayode F.O. (2011). Yields and nutrient pools in soils cultivated with Tectona grandis and Gmelina arborea in Nigerian rainforest ecosystem. Journal of the Saudi Society of Agricultural Sciences, 10:127-135.
- Adelana S.O, Adeosun.T.A, Adesina A.O and Ojuroye M.O. (2011).Environmental pollution and remediation: challenges and management of oil Spillage in the Nigerian coastal areas. Am. J. Sci. Ind. Res., 2011, 2(6): 834-845
- Adhikari K. and Hartemink A.E. (2015). Linking soils to ecosystem services-A global review. Geoderma 262: 101-111.
- Ahmed N. (2016). Application of NDVI in Vegetation Monitoring Using GIS and Remote Sensing in Northern Ethiopian Highlands. Abyss. J. Sci. Technol. Vol. 1, No. 1, 2016, 12-17
- Ahrens D, Schwarzer J, Vogler AP (2014) The evolution of scarab beetles tracks the sequential rise of angiosperms and mammals. Proc R Soc Lond B 281:20141470
- Andreadis, K. M., and Lettenmaier, D. P. (2006). Trends in 20th century drought over the continental United States. Geophys. Res. Lett. 33, L10403, doi:10.1029/2006GL025711.
- Bagchi R, Gallery R.E., Gripenberg S., Gurr S.J., Narayan L., Addis C.E., Freckleton R.P. & Lewis O.T. (2014): Pathogens and insect herbivores drive rainforest plant diversity and composition. Nature 506: 85–88.
- Baker, J.M., Ochsner, T.E., Venterea, R.T., and Griffis, T.J. (2007). Tillage and soil carbon sequestration what do we really know? Agriculture, Ecosystems and Environment 118:1–5.
- Baretta D.; Brown G.G.; James S.W.; Nogueira E.J. and Cardoso B. (2007). Earthworm populations sampled using collection methods in Atlantic Forests with *Araucaria angustifolia* Sci. Agric. (Piracicaba, Braz.); 64(4):384-392
- Barré P., McKenzie B.M. and Hallet P.D., (2009). Earthworms bring compacted and loose soil to a similar mechanical state. *Soil Biol. Biochem.*, 41(3):656-658.
- Baveye PC, Baveye J and Gowdy J (2016) Soil "Ecosystem" Services and Natural Capital: Critical Appraisal of Research on Uncertain Ground. *Front. Environ. Sci.* 4:41. doi: 10.3389/fenvs.2016.00041
- Bernier, N., Ponge, J.F. and Andre. J. (1993). Comparative study of soil organic layers in two bilberry spmce forest stands (Vaccinio-Picetea). Relation to forest dynamics. Geoderma, 59:89-108.
- Bever, J.D., Platt, T.G. and Morton, E.R. (2012) Microbial population and com-munity dynamics on plant roots and their feedbacks on plant communities. Annual Review of Microbiology, 66, 265–283

Vol. 08, No. 03; 2023

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- Bispo, A., Cluzeau, D., Creamer, R., Dombos, M., Graefe, J. U., Krogh, P.H., Soussa, J.P., Peres, G., Rutgers, M., Winding, A. and Römbke, J. (2009). Indicators for monitoring soil biodiversity. *Integrated Environmental Assessment and Management*, 5:717-719.
- Bissett, A., Richardson, A. E., Baker, G. and Thrall, P. H. (2011). Long-term land use effects on soil microbial community structure and function. *Applied Soil Ecology* 51, 66–78,
- Blanchart, E., Albrecht, A., Alegre, J., Duboisset, A., Gilot, C., and Pashanasi, B. (1999). Effects of earthworms on soil structure and physical properties. In: Earthworm Management in Tropical Agroecosystems (eds P. Lavelle, L. Brussaard and P. Hendrix), CAB International, Wallingford, 149–172.
- Blanchart, E., Lavelle, P., Braudeau, E., Le Bissonnais, Y. and Valentin, C. (1997). Regulation of soil structure by geophagous earthworm activities in humid savannas of Cote d'Ivoire. Soil Biology and Biochemistry, 29:431–439.
- Blouin M., Hodson M.E., Delgado E.A., Baker G., Brussaard L., Butt K.R., Dai J., Dendooven L., Peres G., Tondoh J.E., and Brun J.J. (2013). A review of earthworm impact on soil function and ecosystem services. European Journal of Soil Science, 64: 161–182.
- Boeken B, Shachak M, Gutterman Y, and Brand S (1995). Patchiness and disturbance: plant community responses to porcupine diggings in the central Negev. Ecography 18: 410-422
- Bohlen P.J., Parmelee R.W. and Blair J.M. (2004a). Integrating the effects of earthworms on nutrient cycling across spatial and temporal scales. *In*: Edwards C.A., ed. *Earthworm ecology*. 2nd ed. Boca Raton, FL, USA: CRC Press, 183-200.
- Bonanomi, G., Giannino, F. and Mazzoleni, S. (2005). Negative plant-soil feed-back and species coexistence.Oikos,111, 311–321.
- Bonkowski M., Griffiths B.S. and Ritz K. (2000). Food preferences of earthworms for soil fungi. *Pedobiologia*, **44**: 666-676.
- Bossuyt H., Six J. and Hendrix P.F. (2006). Interactive effects of functionally different earthworm species on aggregation and incorporation and decomposition of newly added residue carbon. *Geoderma*, **130** (1-2): 14-25.
- Bouyoucos, G.J. (1926): A calibration of the hydrometer method for making mechanical analysis of soils. *Agronomy Journal*, 43:434 438.
- Brinkman, E.P., Van der Putten, W.H., Bakker, E.J. & Verhoeven, K.J.F.(2010) Plant-soil feedback: experimental approaches, statistical analyses and ecological interpretations. Journal of Ecology,98, 1063–1073
- Brown G.G. and Doube B. (2004). Functional interactions between earthworms, microorganisms, organic matter and plants. Earthworm ecology. 2nd ed. London; Boca Raton, FL, USA: CRC Press, 213-240.
- Brown G.G., Barois I. and Lavelle P. (2000). Regulation of soil organic matter dynamics and microbial activity in the drilosphere and the role of interactions with other edaphic functional domains. *Eur. J. Soil Biol.*, 36(3-4): 177-198.
- Brown, G.G., Pashanasi, B., Gilot-Villenave, C., Patrón, J.C., Senapati, B.K., Giri, S., Barois, I., Lavelle, P., Blanchart. E., Blakemore, R.J., Spain, A.V. & Boyer, J. (1999). Effects of earthworms on plant production. In P. Lavelle, L. Brussaard& P.F. Hendrix, eds. Earthworm management in tropical agroecosystems:87-147. Wallingford, UK, CAB International.
- Brown, M.T, and Buranhan, V. (2003). Energy, indices and ratios for sustainable material cycles and recycle options. Resources, Conservation and Recycling, 38(1): 1-22.

Vol. 08, No. 03; 2023

ISSN: 2456-8643

Bruijnzeel, L.A. (1991). Nutrient input-output budgets of tropical forest ecosystems: A review. Journal of Tropical Ecology, 7:1-24.

Burden, D. S. and Sims, J. L. (1999). Fundamentals of soil science as applicable to management of hazardous wastes. *United States Environmental Protections Agency*, EPA/540/S-98/500.

Butler D. (2006). Virtual globes: the webwide world. Nature 439: 776–78.

- Byers, J.E., Cuddington, K., Jones, C.G., Talley, T.S., Hastings, A., Lambrinos, J.G., Crooks J.A. and Wilson W.G (2006). Using ecosystem engineers to restore ecological systems. *Trends in Ecology & Evolution*, 21:493–500.
- Chandra, L.R., Gupta, S., Pande, V. *et al.* Impact of forest vegetation on soil characteristics: a correlation between soil biological and physico-chemical properties. *3 Biotech* 6, 188 (2016). <u>https://doi.org/10.1007/s13205-016-0510-y</u>
- Condit R., Engelbrecht B.M.J., Pino D., Pe´rez R., and Turner B.L. (2013). Species distributions in response to individual soil nutrients and seasonal drought across a community of tropical trees. Proc Nat Acad Sci 110:5064–5068
- Chima U.D. and Omokhua G.E. (2011): Vegetation Assessment and Description. In Aiyeloja A.A. and Ijeomah H.M (Eds): Book of reading in Forestry, Wildlife Management and Fisheries. 104-129.
- Cook, T. D., and Campbell, D. T. (1979). *Quasi-experimentation: Design & analysis issues in field settings*. Boston, MA: Houghton Mifflin
- Corlett R.T. and Primack R.B. (2006): Tropical rainforests and the need for cross-continental comparisons. Trends Ecol Evol 21:104–110
- Cunha, L., Brown G.G., Stanton, D, Da Silva E, Hausel F, Jorge G, Mckey D, Vidal-Toroado P, Macedo R, Velasquez E., James S, Lavelle P, and Kille P. (2006). Soil Animals and Pedogenesis. The role of earthworms in Anthopogenic soils. Soil Science, 181:110 125.
- Cyme (1999). Soil microbiology an exploration approach, Delma.
- Dash M.C. and Patra U.C., (1979). Wormcast production and nitrogen contribution to soil by a tropical earthworm population from a grassland site in Orissa, India. *Rev. Ecol. Biol. Sol*, 16(1): 79-83.
- Davies R.G., Eggleton P., Jones D.T., Gathorne-Hardy F.J., and Hernandez L.M. (2003). Evolution of termite functional diversity: analysis and synthesis of local ecological and regional influences on local species richness. J Biogeogr 30:847–877.
- De Deyn G.B, Quirk H, Yi Z, Oakley S, Ostle N.J. and Bardgett R.D. (2009). Vegetation composition promotes carbon and nitrogen storage in model grassland communities of contrasting soil fertility. J Ecol., 97:864–75. doi: 10.1111/j.1365-2745.2009.01536.x.
- Decaëns, T.; Jiménez, J.J.; Barros, E.; Chauvel, A.; Blanchart, E.; Fragoso, C. and Lavelle, P. (2004). Soil macrofauna communities in permanent pastures derived from tropical forest or savanna. Agriculture, Ecosystems and Environment 103: 301-312.
- Drake H.L. and Horn M.A. (2007). As the worm turns: the earthworm gut as a transient habitat for soil microbial biomes. *Annu. Rev. Microbiol.*, 61:169-189.
- Drenovsky, R. E., Steenwerth, K. L., Jackson, L. E. and Scow, K. M. (2010). Land Use and Climatic Factors Structure Regional Patterns in Soil Microbial Communities. *Global Ecology and Biogeography* 19, 27–39,
- Dutta, R.K. and Agrawal, M. (2002). Effect of tree plantations on the soil characteristics and microbial activity of coal mine spoil land. Tropical Ecology 43(2): 315-324.

Vol. 08, No. 03; 2023

ISSN: 2456-8643

- Edwards C.A. and Bohlen P.J., (1996). Earthworm ecology and biology. London: Chapman & Hall, 196-212
- Emaziye, P. O., Okoh R. N. and Ike P. C. (2012). "A Critical Analysis of Climate Change Factors and its Projected Future Values in Delta State, Nigeria", Asian Journal of Agriculture and Rural Development, 2(2):206-212.
- Ernst G. and Emmerling C., (2009). Impact of five different tillage systems on soil organic carbon content and the density, biomass and community composition of earthworms after a ten year period. *Eur. J. Soil Biol.*, 45(3):247-251.
- Ettema, C. H., & Wardle, D. A. (2002). Spatial soil ecology. Trends in Ecology and Evolution, 17:177-183.
- Eze A.G. (2015). Relevance of the Tort of Nuisance in Redressing Damage From Oil and Gas Pollution In Nigeria. NAUJILJ 2015: 147-155
- Felske A., Akkermans A.D.L. and De Vos W.M. (1998). *In situ* detection of an uncultured predominant *Bacillus* in Dutch grassland soils. *Appl. Environ. Microbiol.*, 64(11):4588-4590.
- Fitter, A.H., Gilligan, C. A., Hollingworth, K., Kleczkowski, A., Twyman, R.M., Pitchford J. W. and The Members of the NERC Soil Biodiversity Programme (2005): Biodiversity and ecosystem function in soil. Functional Ecology, 19:369-377
- Foissner W. (2006): Biogeography and dispersal of micro-organisms:a review emphasizing protists. Act Protozool 45:111–136
- Fonte S.J., Thaiis W., and Six J. (2009). Earthworm populations in relation to soil organic matter dynamics and management in California tomato cropping systems. European Journal of Soil Biology, 4: 206–214
- Fragoso, C. and Lavelle, P. (1992). Earthworm communities of tropical rain forests. Soil Biol. Biochem. 24:1397–1408.
- Freitag A., Rudert M. and Bock E. (1987). Growth of *Nitrobacter* by dissimilatoric nitrate reduction. *FEMS Microbiol. Lett.*, 48(1-2):105-109.
- Fujii K, Hartono A, Funakawa S, Uemura M, Sukartiningsih Kosaki T (2011): Distribution of Ultisols and Oxisols in the serpentine areas of East Kalimantan, Indonesia. Pedologist 55:63–76.
- Fujii K. (2014) Soil acidification and adaptations of plants and microorganisms in Bornean tropical forests. Ecol Res. 29:371–381
- Fujii K., Shibata M., Kitajima K., Ichie K., Kitayama K. & Turner B.L. (2018). Plant-soil interactions maintain biodiversity and functions of tropical forest ecosystems. Ecol Res (2018) 33: 149–160. DOI 10.1007/s11284-017-1511-y
- Fukami T. and Nakajima M. (2013). Plant–Soil Feedbacks in A Changing World Complex plant–soil interactions enhance plant species diversity by delaying community convergence. Journal of Ecology, 101, 316–324.
- Glaser B. & Birk J.J. (2012). State of the scientific knowledge on properties and genesis of Anthropogenic Dark Earths in Central Amazonia (terra preta de I' ndio). Geochimica et Cosmochimica Acta, 82:39–51
- Godfray, C.H.J., Beddington, J.R., Crute, I.R., Haddad, L., Lawrence, D., Muir, J.F., Pretty, J., Robinson, S., Thomas, S.M. and Toulmin, C. (2010). Food Security: The Challenge of Feeding 9 Billion People. *Science*, 327, 812-818.

Vol. 08, No. 03; 2023

ISSN: 2456-8643

- Guo, X. (2016). Effects of land use change on the composition of soil microbial communities in a managed subtropical forest. *Forest Ecology and Management* 373, 93–99.
- Hayakawa C, Funakawa S, Fujii K, Kadono A, and Kosaki T (2013).Effects of climatic and soil properties on cellulose decomposition rates in temperate and tropical forests. Biol Fertil Soil50:633–643
- Heineman K.D., Turner B.L., and Dalling J.W. (2016): Variation in wood nutrients along a tropical soil fertility gradient. New Phytol 211:440–454
- Hidaka A, and Kitayama K. (2009) Divergent patterns of photosynthetic phosphorus-use efficiency versus nitrogen-use efficiency of tree leaves along nutrient-availability gradients. J Ecol 97:984–991
- Ichikogu, V.I. (2012). Total nitrogen and available phosphorus dynamics in soils regenerating from degraded abandoned rubber plantation in Orogun Area of the rainforest zone of southern Nigeria. Ethiopian Journal of Environmental Studies and Management, 5(1):92-99.
- Ihssen J., Horn M.A., Matthies C., Gobner A., Schrammand A, and Drake H.L. (2003). N₂O producing microorganisms in the gut of the earthworm *Aporrectodea caliginosa* are indicative of ingested soil bacteria. *Appl. Environ. Microbiol.*, 69(3): 1655-1661.4-131.
- Kardol, P., Cornips, N.J., van Kempen, M.M.L., Bakx-Schotman, J.M.T. and vander Putten, W.H. (2007) Microbe-mediated plant-soil feedback causes histori-cal contingency effects in plant community assembly. Ecological Mono-graphs, 77, 147–162.
- Karnieli A., Agam N, Pinker R.T., Anderson M., Imhoff M.I., Gutman G.G., and Panov N. (2010). Use of NDVI and Land Surface Temperature for Drought Assessment: Merits and Limitations. Journal of Climate, 23(3):618-633.
- . Proceedings of the Royal Society B-Biological Sciences, 279, 3020–3026.
- Lal, R. (2011). Soil health and climate change: An overview. In Soil Health and Climate Change (Eds.,Singh, B. P., Cowie, A. L. and Chan, K. Y.), Soil Biology Series (29):3–24. Springer-Verlag, Berlin.
- Landesman W and Dighton J (2011): Shifts in microbial biomass and the bacteria: fungi ratio occur under field conditions within 3 h after rainfall. Microb Ecol 62:228–236. doi:10.1007/s00248-011-9811-1
- Lemtiri A., Colinet G., Alabi T, Cluzeau D., Zirbes L., Haubruge E., and Francis F. (2014). Impacts of earthworms on soil components and dynamics. A review. Biotechnol. Agron. Soc. Environ. 2014 18(1): 1-13
- Li X., Fisk M.C., Fahey T.J. and Bohlen P.J. (2002). Influence of earthworm invasion on soil microbial biomass activity in a northern hardwood forest. *Soil Biol. Biochem.*, 34(12):1929-1937.
- Ologunorisa T.E. and Adejuwon J.O. (2003). Annual rainfall trends and periodicities in the Niger Delta, Nigeria. *Journal of Meteorology* 28(276): 41–51.
- Owa S.O., Dedeke G.A., Morafa S.O.A. and Yeye J.A. (2003). Abundance of earthworms in Nigerian ecological zones: implications for sustaining fertilizer-free soil fertility. African Zoology, 38(2):235-244.
- Rousk J, Baath E, Brookes P.C., Lauber C.L., Lozupone C, Caporaso J.G., Knight R, and Fierer N (2010) Soil bacterial and fungal communities across a pH gradient in an arable soil. ISME4:1340–1351

Vol. 08, No. 03; 2023

ISSN: 2456-8643

- Shen H. (2011). Land Use Spatial Pattern Characteristics along the Terrain Gradient in Yellow River Basin in West Henan Province, China. IEEE; 1-5.
- Sollins P (1998): Factors influencing species composition in tropical lowland rain forest: does soil matter? Ecology 79:23–30.
- Umeuduji, J. E and Aisuebeogun, A., (1999). Relief and Drainage. In C. U. Oyegun and A. Adeyemo, (eds.): Port Harcourt Region, Port Harcourt: Department of Geography and Environmental Management, Publication Series; No.1
- Vicent, S., Pons, F.N., and Cuadrat, P. (2004). 'Mapping soil moisture in the central Ebro river valley (Northeast Spain) with Landsat and NOAA satellite imagery: a comparison with meteorological data'. International Journal ofRemote Sensing., 25: 4325-4350.
- Vitousek P.M. (2004) Nutrient cycling and limitation: Hawai'i as a model system. Princeton University Press, Princeton
- Walkey, A. and Black, C.A. (1934). "An examination of the dejareff method of determining soil organic matter and a proposed modification of chromic acid titration method", *Soil Science*, 37:29-38.
- Wallis, P., Haynes, R., Hunter, C. and Morris, C. (2010). Effect of land use and management on soil bacterial biodiversity as measured by PCR-DGGE. Applied Soil Ecology 46, 147–150
- Wardle (2004). Ecological linkages between aboveground and belowground biota. Science, 304:1631-1633.